

ORIGINAL RESEARCH

Staphylococcus aureus-derived lipoteichoic acid induces inflammation and alters skin barrier function in atopic dermatitis

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ABSTRACT

Background: The present study was conducted for assessing alteration of skin barrier function by *Staphylococcus aureus*-derived lipoteichoic acid (ALA) in atopic dermatitis. **Material and methods:** For evaluating alteration of skin barrier function by *Staphylococcus aureus*-derived lipoteichoic acid in atopic dermatitis patients, a total of 20 patients were evaluated. Inclusion criteria for the present study included patients with presence of atopic dermatitis. In-vitro samples were obtained and direct exposure of T cells was done. Fluorescein isothiocyanate contact hypersensitivity models for TH2-mediated cutaneous inflammation. All the results were recorded in Microsoft excel sheet and were subjected to statistical analysis using SPSS software. **Results:** ALA was found to effectively inhibit the activation of T lymphocytes in a manner that does not depend on Toll-like receptor 2. T cells that were exposed to ALA exhibited neither proliferation nor cytokine production. Consequently, exposure to ALA led to a transient state of functional paralysis in T cells. Furthermore, ALA significantly diminished both T-cell cytokine production and cutaneous recall responses. **Conclusion:** Improved identification and characterization of atopic dermatitis is required to optimize the precision medicine approach.

Keywords: *Staphylococcus*, Lipoteichoic acid

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INTRODUCTION

Staphylococcus aureus is a gram-positive extracellular bacterium responsible for a diverse array of human diseases, leading to nearly 500,000 hospital admissions annually in the United States.^{1,2} Infections typically initiate in the skin, where epidermal keratinocytes play a crucial role as the initial defense mechanism against bacterial invasion as part of the innate immune response. Previous investigations into the keratinocyte response to various *S. aureus* components revealed that cell wall lipoteichoic acid (LTA) significantly altered gene expression patterns.¹³ Notably, a gene array analysis conducted on primary keratinocytes indicated that LTA influenced the regulation of over 300 genes. Specifically, it was observed that LTA downregulated several genes

associated with keratinocyte differentiation, a process that was found to be dependent on the transcription factor p63, which is essential for keratinocyte proliferation and skin development.⁴⁻⁶

Conversely, the in vivo effects of LTA on skin remain largely unexamined. While *S. aureus* LTA is absent from healthy skin, physiological levels have been identified in the lesional skin of many patients suffering from atopic dermatitis (AD)⁷, a condition characterized by an inflammatory skin response, a compromised skin barrier, and diminished expression of vital barrier proteins such as filaggrin and loricrin.⁸⁻¹⁰

Atopic dermatitis (AD) is a chronic pruritic inflammatory skin disease, whose pathogenesis is mediated by interactions between skin barrier

impairment and an abnormal immune response featuring enhanced type 2 inflammation. Interactions between keratinocytes (KCs), innate immune cells (e.g., type 2 innate lymphoid cells [ILC2s], dendritic cells, mast cells, basophils, and eosinophils), adaptive immune cells (T and B cells), and an altered epidermal microbiome (with reduction of microbial diversity and predominance of *Staphylococcus aureus*) all contribute to AD pathogenesis.¹¹⁻¹³ Hence; the present study was conducted for assessing alteration of skin barrier function by *Staphylococcus aureus*-derived lipoteichoic acid in atopic dermatitis.

MATERIAL AND METHODS

For evaluating alteration of skin barrier function by *Staphylococcus aureus*-derived lipoteichoic acid in atopic dermatitis patients, a total of 20 patients were evaluated. Inclusion criteria for the present study included patients with presence of atopic dermatitis. In-vitro samples were obtained and direct exposure of T cells was done. Fluorescein isothiocyanate contact hypersensitivity models for TH2-mediated cutaneous inflammation. All the results were recorded in Microsoft excel sheet and were subjected to statistical analysis using SPSS software.

RESULTS

ALA was found to effectively inhibit the activation of T lymphocytes in a manner that does not depend on Toll-like receptor 2. T cells that were exposed to ALA exhibited neither proliferation nor cytokine production. Consequently, exposure to ALA led to a transient state of functional paralysis in T cells. Furthermore, ALA significantly diminished both T-cell cytokine production and cutaneous recall responses.

DISCUSSION

The microbial community, referred to as the "microbiome," plays a dual role, exhibiting both advantageous and harmful effects. For instance, *Staphylococcus epidermidis*, a major inhabitant of healthy human skin, has been shown to mitigate inflammation following skin injuries, uphold immune tolerance towards commensal organisms, influence the development of cutaneous T-cells, and bolster innate immune responses by promoting the expression of antimicrobial peptides (AMPs).¹¹⁻¹⁵

In contrast, an imbalance in the microbiome, known as dysbiosis, has been implicated in the development of various skin disorders. Notably, a strong correlation exists between dysbiosis and the clinical manifestations of atopic dermatitis (AD).¹⁶ Individuals with AD are particularly characterized by heightened colonization of *Staphylococcus aureus* (*S. aureus*) and a reduction in skin bacterial diversity. Moreover, recent mechanistic investigations have revealed that *S. aureus* can induce the formation of AD-like lesions in murine models. These observations indicate that a deeper comprehension of the

interactions between bacteria and skin immunity could yield significant insights for enhancing the management of AD.¹⁷⁻²⁰

ALA was found to effectively inhibit the activation of T lymphocytes in a manner that does not depend on Toll-like receptor 2. T cells that were exposed to ALA exhibited neither proliferation nor cytokine production. Consequently, exposure to ALA led to a transient state of functional paralysis in T cells. Furthermore, ALA significantly diminished both T-cell cytokine production and cutaneous recall responses. *Staphylococcus aureus* is capable of provoking inflammation through mechanisms such as the promotion of T cell-independent B cell proliferation, which leads to the secretion of proinflammatory cytokines like thymic stromal lymphopoietin (TSLP) from keratinocytes, as well as the activation of mast cell degranulation, ultimately resulting in a TH2 skewing response.²¹⁻²⁴ Additionally, *S. aureus* alters the proteolytic equilibrium within the skin by stimulating various metalloproteases in dermal fibroblasts.²⁵

Nevertheless, due to the intricate architecture and cellular networks present in mammalian skin, the precise mechanisms through which *S. aureus* disrupts cutaneous inflammatory homeostasis remain inadequately elucidated. It seems that the beneficial and harmful effects of skin-associated bacteria largely hinge on their ability to engage with host cells located beneath the stratum corneum. Until recently, the pathways through which microbes residing on the skin surface could modulate immune responses via the stratum corneum structure were not well understood.²⁶

CONCLUSION

Improved identification and characterization of atopic dermatitis is required to optimize the precision medicine approach.

REFERENCES

1. Barton CE, Johnson KN, Mays DM, Boehnke K, Shyr Y, Boukamp P, et al. Novel p63 target genes involved in paracrine signaling and keratinocyte differentiation. *Cell Death Dis* 2010;1:e74.
2. Boguniewicz M, Leung DY. Atopic dermatitis: a disease of altered skin barrier and immune dysregulation. *Immunol Rev* 2011;242(1):233–46.
3. Brauweiler AM, Goleva E, Hall CF, Leung DYM. Th2 Cytokines Suppress Lipoteichoic Acid-Induced Matrix Metalloproteinase Expression and Keratinocyte Migration in Response to Wounding. *J Invest Dermatol* 2015;135(10):2550–3.
4. Brauweiler AM, Hall CF, Goleva E, Leung DYM. *Staphylococcus aureus* Lipoteichoic Acid Inhibits Keratinocyte Differentiation through a p63-Mediated Pathway. *J Invest Dermatol* 2017;137(9):2030–3.
5. Choy DF, Hsu DK, Seshasayee D, Fung MA, Modrusan Z, Martin F, et al. Comparative transcriptomic analyses of atopic dermatitis and psoriasis reveal shared neutrophilic inflammation. *J Allergy Clin Immunol* 2012;130(6):1335–43 e5.

6. Dhingra N, Suarez-Farinas M, Fuentes-Duculan J, Gittler JK, Shemer A, Raz A, et al. Attenuated neutrophil axis in atopic dermatitis compared to psoriasis reflects TH17 pathway differences between these diseases. *J Allergy Clin Immunol* 2013;132(2):498–501 e3.
7. Foster SL, Hargreaves DC, Medzhitov R. Gene-specific control of inflammation by TLR-induced chromatin modifications. *Nature* 2007;447(7147):972–8.
8. Guttman-Yassky E, Nograles KE, Krueger JG. Contrasting pathogenesis of atopic dermatitis and psoriasis—part I: clinical and pathologic concepts. *J Allergy Clin Immunol* 2011;127(5):1110–8.
9. Hersh AL, Chambers HF, Maselli JH, Gonzales R. National trends in ambulatory visits and antibiotic prescribing for skin and soft-tissue infections. *Arch Intern Med* 2008;168(14):1585–91.
10. Iwamoto K, Numm TJ, Koch S, Herrmann N, Leib N, Bieber T. Langerhans and inflammatory dendritic epidermal cells in atopic dermatitis are tolerized toward TLR2 activation. *Allergy* 2018;73(11):2205–13.
11. Akaiwa M., Yu B., Umeshita-Suyama R., Terada N., Suto H., Koga T., et al. Localization of human interleukin 13 receptor in non-haematopoietic cells. *Cytokine*. 2001;13:75–84.
12. Akdis C.A. Does the epithelial barrier hypothesis explain the increase in allergy, autoimmunity and other chronic conditions? *Nat Rev Immunol*. 2021;21:739–751.
13. Amano W., Nakajima S., Kunugi H., Numata Y., Kitoh A., Egawa G., et al. The Janus kinase inhibitor JTE-052 improves skin barrier function through suppressing signal transducer and activator of transcription 3 signaling. *J Allergy Clin Immunol*. 2015;136:667–677.e7.
14. Bekeredjian-Ding I, Inamura S, Giese T, Moll H, Endres S, Sing A, et al. Staphylococcus aureus protein A triggers T cell-independent B cell proliferation by sensitizing B cells for TLR2 ligands. *J Immunol*. 2007;178:2803–12.
15. Bisgaard H, Simpson A, Palmer CN, Bonnelykke K, McLean I, Mukhopadhyay S, et al. Gene-environment interaction in the onset of eczema in infancy: filaggrin loss-of-function mutations enhanced by neonatal cat exposure. *PLoS Med*. 2008;5:e131.
16. Camargo CA, Jr, Ganmaa D, Sidbury R, Erdenedelger K, Radnaakhand N, Khandsuren B. Randomized trial of vitamin D supplementation for winter-related atopic dermatitis in children. *J Allergy Clin Immunol*. 2014;134:831–5 e1.
17. Cogen AL, Yamasaki K, Sanchez KM, Dorschner RA, Lai Y, MacLeod DT, et al. Selective antimicrobial action is provided by phenol-soluble modulins derived from Staphylococcus epidermidis, a normal resident of the skin. *J Invest Dermatol*. 2010;130:192–200.
18. Dancer SJ, Garratt R, Saldanha J, Jhoti H, Evans R. The epidermolytic toxins are serine proteases. *FEBS Lett*. 1990;268:129–32.
19. De Benedetto A, Rafaels NM, McGirt LY, Ivanov AI, Georas SN, Cheadle C, et al. Tight junction defects in patients with atopic dermatitis. *J Allergy Clin Immunol*. 2011;127:773–86. e1–7.
20. Di Nardo A, Yamasaki K, Dorschner RA, Lai Y, Gallo RL. Mast cell cathelicidin antimicrobial peptide prevents invasive group A Streptococcus infection of the skin. *J Immunol*. 2008;180:7565–73.
21. Dorschner RA, Pestonjamas VK, Tamakuwala S, Ohtake T, Rudisill J, Nizet V, et al. Cutaneous injury induces the release of cathelicidin anti-microbial peptides active against group A Streptococcus. *J Invest Dermatol*. 2001;117:91–7.
22. Fallon PG, Sasaki T, Sandilands A, Campbell LE, Saunders SP, Mangan NE, et al. A homozygous frameshift mutation in the mouse Flg gene facilitates enhanced percutaneous allergen priming. *Nat Genet*. 2009;41:602–8.
23. Foelster Holst R, Reitamo S, Yankova R, Worm M, Kadurina M, Thaci D, et al. The novel protease inhibitor SRD441 ointment is not effective in the treatment of adult subjects with atopic dermatitis: results of a randomized, vehicle-controlled study. *Allergy*. 2010;65:1594–9.
24. Gallo RL, Hooper LV. Epithelial antimicrobial defence of the skin and intestine. *Nat Rev Immunol*. 2012;12:503–16.
25. Gallo RL, Nakatsuji T. Microbial symbiosis with the innate immune defense system of the skin. *J Invest Dermatol*. 2011;131:1974–80.
26. Gallo RL, Ono M, Povsic T, Page C, Eriksson E, Klagsbrun M, et al. Syndecans, cell surface heparan sulfate proteoglycans, are induced by a proline-rich antimicrobial peptide from wounds. *Proc Natl Acad Sci U S A*. 1994;91:11035–9.